New resources for field crop disease management

Kaitlyn Bissonnette

Much like the development of herbicide-resistant weeds, repeated use of chemical control methods can result in the selection of fungicide-resistant pathogen isolates in the field. In an effort to preserve the effectiveness of herbicide technology, the Take Action pesticide resistance program was launched in 2013 as Take Action on Weeds. This January, it has expanded to include fungicide resistance management as part of its farmer-focused educational platform as Take Action on Diseases. The program website includes resources on fungicide resistance management for field crops, including a Fungicide Classification poster.

In addition to the Take Action program, other resources are available with information on fungicide resistance management (Bradley et al. 2014 and CPN-4001) and fungicide efficacy for field crops. Fungicide efficacy tables are constructed each year from data gathered by independent, unbiased University researchers across the United States. Tables are available for foliar diseases of corn, soybean, and small grains and seedling diseases of soybean. The most commonly used products are tested in these trials, so the list is not comprehensive of all products that are commercially available. When using fungicides, follow all label recommendations on the container. The label is the law! Keep in mind that fungicides are a tool, not a cure. Integrated pest management practices are necessary to preserve their effectiveness.

The Crop Protection Network (CPN) also has re-vamped its webpage and includes a whole host of field crop disease resources compiled by university, government, and industry experts which are reviewed and unbiased. Fact sheets, videos, and live twitter feeds from CPN experts are all readily available on the CPN website. Resources are currently available for corn, soybeans, and small grains, including disease loss estimates, disease fact sheets, scouting resources, and fungicide FAQ sheets.

More information on Take Action on Diseases can be found at http://iwilltakeaction.com/diseases and on the Crop Protection Network and its resources can be found at https://cropprotectionnetwork.org.
Mizzou Integrated Pest Management Field Day - July 10

Mandy D. Bish

The annual Mizzou Integrated Pest Management Field Day will be Tuesday July 10th at the Bradford Research and Extension Center, approximately 8 miles east of Columbia, Missouri. Meet new extension state pathologist Kaitlyn Bissonnette, new state extension entomologist Kevin Rice, get the latest updates on dicamba, and take the guided wagon tours with stops that feature the latest research. Details on registration will be posted in June.

To learn more about Mizzou Weed Science, visit the Web site at www.weedscience.missouri.edu or find us on Facebook and Twitter at Mizzou Weed Science.

Reopening of the Plant Diagnostic Clinic 2018

Josephine Mgbechi-Ezeri

Hurray! The Plant Diagnostic Clinic is back again. The clinic was established to provide answers to plant health problems faced by the citizens of Missouri. The mission of the clinic is to provide accurate, timely answers and management recommendation that reflects research-based results and an integrated pest management (IPM) philosophy. Besides answering plant disease problems, the clinic also handles samples submitted for identification of weeds, mushroom, insects or arachnids. Starting January 8th, Josephine Mgbechi-Ezeri will serve as the new clinic Director. Josephine’s role is to coordinate the daily operations of the laboratory, provide diagnostic services to clients and disease management training to the agricultural stakeholders in Missouri. When necessary, the clinic can utilize the expertise of University of Missouri State Extension Specialists and faculty in the Division of Plant Sciences who specialize in Agronomy, Entomology, Horticulture or Plant Pathology to ensure accurate and effective diagnosis and reporting. We welcome samples from government agencies, growers, industries and homeowners throughout the state.

Please visit the Plant Diagnostic Clinic website for:

- Information on how to collect and ship a sample
- Submission forms (types of samples accepted)
- Plant Disease Identification
- Turfgrass Disease Identification
- Insect / Arachnid Identification
- Plant / Weed Identification (to include mushrooms)
- Clinic hours for dropping off a sample
- Fees associated with services

Contact information:
University of Missouri–Plant Diagnostic Clinic
28 Mumford Hall
Columbia, MO 65211

Phone: 573-882-3019
Email: plantclinic@missouri.edu
Web: http://plantclinic.missouri.edu/
Introduction

The invasive Japanese beetle, Popillia japonica, poses a pest management challenge for crop farmers, in particular organic producers. Considering that organic options for the management of this pest are limited, developing a mass trapping system to control this pest is a relatively new approach. At first, traps baited with the Japanese beetle sex pheromone were created for the purpose of monitoring. In this article we present results from research conducted from 2012 to 2017 by the Lincoln University Integrated Pest Management (IPM) program, which aimed at developing a mass trapping system that could provide effective Japanese beetle control in agricultural areas with less or no insecticides applied to the crop. Both organic and non-organic farmers may find this information useful. We also provide detailed instructions on how to develop traps and how to get the lures.

Seasonal Activity of Japanese Beetles.

In mid-Missouri, significant numbers of Japanese beetles begin emerging in mid to late June, the population usually peaks the second week of July, and declines by early August. The timing of the onset and end of Japanese beetle adult activity may vary by a couple weeks depending on weather. For instance, if warm, humid conditions occur—they will emerge earlier. If it’s cold and rainy, they may not become active until late June.

What is Mass Trapping?

Mass trapping is a behaviorally-based method of reducing pest numbers by luring them in large numbers to a trap or device that contains an attractant (usually a food component or a pheromone, in some cases in combination with attractive colors) and then killing the pests either, with a toxicant or a mechanism that prevents them from leaving the traps. For Japanese beetles, the two main trap designs that have been evaluated are presented below.


For research purposes, a mass trapping design that consisted of an aluminum mesh sock 4 feet long by one foot in diameter was evaluated in most years. Velcro was used to secure the sock to the yellow funnel with duct tape for added strength. Seams were stapled. Research conducted during 2015 and 2016 compared the effectiveness of 32 gallon capacity black plastic bins that require less maintenance to that of the aluminum mesh sock. For detailed instructions on how to construct mass trapping devices, refer to our guide on building mass trapping systems for Japanese Beetles on the back cover.

All traps are baited with a double lure system comprised of a floral-based lure and the Japanese beetle sex pheromone.
These lures are for agricultural use. Japanese beetle lures are always used in conjunction with trap tops that consist of yellow panels that intersect at 90° with a funnel underneath ending in a wide rim. Beetles hitting the vane fall through the funnel into the collecting device. Yellow tops and lures can be purchased from Great Lakes IPM: http://www.greatlakesipm.com.

Prior to deploying the mass trapping system it is recommended to hang a single monitoring trap in late May in mid-Missouri. Check it regularly so you know exactly when the pest arrives and deploy the full spread of traps to make a “force field” once the first beetles are captured in the monitoring trap. Place traps along the entire perimeter—like a fence—around the crop you wish to protect. Traps are meant to be a barrier to intercept beetles before they land on crops. However, if the area is too big or the number of traps is too small for that, you can place them along the side(s) of highest pressure. Japanese beetle larvae feed on the roots of grass. This makes fescue pasture an ideal breeding ground. If there is a large area of grass (backyard, golf course, or pasture), that is likely to be the side of highest Japanese beetle pressure. Always consider placing traps upwind. Do not place traps in the center of the field because that will make things worse. Once Japanese beetles are happily feeding and mating on plants, they will be reluctant to leave unless shaken off the foliage, pushed away by a repellent (such as kaolin clay “Surround”), or by spraying an OMRI-listed insecticide.

 Trap Deployment Patterns.

The “ideal” number of traps to be used for a given area depends on the size of the plot, pest pressure, and time / resources available. From 2012 to 2017 we evaluated the ability of novel mass trapping systems to capture Japanese beetles in elderberry and blueberry orchards located at two Missouri locations. The first location was a 0.5 acre elderberry orchard located at the Lincoln University George Washington Carver farm in Jefferson City, Missouri. This orchard comprises nine elderberry genotypes: Bob Gordon, Dallas, Deer, Marge, Ocoee, Ozone, Sperandio, Wyldewood, and York. The second location was a 2.5 acre blueberry orchard at the Lincoln University Alan T. Busby organic research farm. This orchard comprises three blueberry cultivars (Duke, Liberty, and Blue Crop).

For blueberries, 16 traps have proven to be effective at suppressing beetles from the blueberry plants. In our studies, traps have been deployed along two of the four orchard sides based on the direction of Japanese beetle pressure, with about 20 yards between traps. Since elderberries are more attractive to Japanese beetles than blueberry, at the elderberry farm traps have been spaced about 5 yards apart. As you place the traps around the perimeter, be sure to leave a buffer zone of approximately 10 yards between the traps and the crop. Do not put traps too close to the crop because you don’t want residual beetles swarming around the traps to accidently land on foliage.

**Main Research Findings.**

**Beetle Densities on Crop Plants and Level of Feeding Damage to Crop Foliage**

Visual inspections of Japanese beetle feeding damage to perimeter row plants were performed at both farms. The number of beetles per plant and the level of defoliation to the nearest 5th percentile (0-5%, 6-10%, 11-15%) were recorded. At Carver farm, the entire perimeter (60 plants) was checked on a weekly basis. At Busby farm, 100 blueberry plants in the perimeter row and 100 plants in row 3 were checked on a weekly basis. Only data from 2014 to 2016 are presented below.

At the elderberry orchard, the season-long average number of Japanese beetles was only 0.5, 3.7, and 1.9 adult Japanese beetles per plant, in 2014, 2015, and 2016, respectively. The season-long mean percent defoliation was 2.5% in 2014, 8.2% in 2015, and 9.7% in 2016 on average. Thus, the number on elderberries remained below economic threshold (<5 beetles per plant) and defoliation was less than 10% for all 3 years while traps succeeded in catching massive number of adults.

At the blueberry orchard, the season-long average number of beetles on foliage was 0.06, 0.07, and 0.01 per plant, for 2014, 2015, and 2016 respectively. The season-long mean percent defoliation was, on average, 0.3% in 2014, 0.07% in 2015, and 0.02% in 2016. Visual inspections
that negative impacts of insecticide application on non-targets will be avoided.

If a person sprays PyGanic twice a week for six weeks then the season-long cost estimate is for the cost of insecticide only, without including time savings, etc. Such estimate is for the cost of insecticide only, without including time savings, etc. If a person sprays PyGanic twice a week for six weeks then the season-long cost of spraying organic insecticides would amount to $924.

Costs

Estimated costs associated with the construction of one mass trapping device using a trash bin is $30.50 (approx. cost of yellow top: $10.50; cost of bin with lid: $15.00; cost of mesh + glue: $5.00). Assuming deployment of 7 trash bins per acre (general trap density used at the blueberry orchard), then the total cost of traps will be: $213.50 (a one-time investment). Accordingly, the annual cost of 14 lures ($4.25 each), including one replacement (done during the peak of Japanese beetle activity) is $59.50.

In contrast, the cost of spraying PyGanic 5.0 EC against Japanese beetle in one-acre plot is approx. $77.00 per application, using the high label rate. Such estimate is for the cost of insecticide only, without including time savings, etc. If a person sprays PyGanic twice a week for six weeks then the season-long cost of spraying organic insecticides would amount to $924. Use of traps also means that negative impacts of insecticide application on non-targets will be avoided.

Advantages of Using a Large Capacity Mass Trapping System

By using large trash bins, traps may not need to be emptied once a week. If the bins need to be emptied, then the easiest way to do this is to raise the lid slightly, slide a garbage bag over the opening, and pull it down so that no beetles can escape. Dump the bin upside down so the plastic bag is on the ground and smack the bottom of the bin with the palm of your hand. The beetles will fall into the garbage bag effortlessly. You can easily dump the beetles from one plastic bag into another until it's full. Any live beetles will suffocate quickly in the sealed plastic bag. Once all beetles are dead, you can tear the bag open, dump the dead beetles on a compost pile, and throw the plastic bag away.

While it takes a bit of time to empty and maintain the traps, this would be offset by the time and insecticide cost savings in not having to spray a crop.

(continued on back page)

References:


To avoid having to empty the trash bins, beetles can be composted on-site, by adding carbon sources (wooden mulch, dry leaves, moisten shredded paper) periodically to form layers of dead beetles and carbon sources. Proper composting can help minimize or avoid the foul smell of dead Japanese beetles, which is caused mostly by release of ammonia.

**How to Construct a Large-Capacity Mass Trapping System**

**Step 1**: Drill small holes in bottom of the 32 gallon bin with 3/8ths inch bit to allow rain water to drain.

**Step 2**: Cut a 5 by 15 inch rectangle out of cardboard. Use as a template to draw 2 windows (one on each side of bin).

**Step 3**: Use an aviation “tin-snip” tool to cut out the window you traced with permanent marker.

**Step 4**: The front and back of bin should look like this.

**Step 5**: Stand the bin on it’s base.

**Step 6**: Use sandpaper to sand the inside edges of the windows. This will allow the glue to adhere to the plastic. WARNING: Do not use hot glue because it will melt outside in the summer heat!

**Step 7**: Cut a 7 by 20 inch rectangle out of cardboard. Use this template to cut out 2 window screens.

**Step 8**: Lay the bin on one side and secure the aluminum screen inside with duct tape. Glue the screen to the bin and allow glue to dry overnight. Gorilla glue is best—you can also use epoxy or E6000.

**Step 9**: Both steel mesh windows in place.

**Step 10**: At the center of the lid, draw an “X” that is 4 inches by 4 inches. Draw a 2 inch circle around the center.

**Step 11**: Drill a large hole in the center and use that as a starting point to cut out the “X” & circle shape you drew.

**Step 12**: It should look like this.

**Step 13**: Push the base of yellow commercial top through the slots you made.

**Step 14**: Peel the silver tab off of lure, remove rubber pheromone stopper, and place in cutout hole.

**Step 15**: Use Gorilla tape or duct tape to seal the cracks so beetles can’t escape.

**Step 16**: Snap lid onto bin. To keep the bin upright, we used two 4 ft. metal "electric fence posts" slid through the handles (price: about $1.50 each). You can also use Rebar or simply hammer a tomato stake in the ground. Then, wrap a piece of string or wire around the bin to secure it.